

## Application Note K2 Transfection System

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### Experimental Details

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**Cell type:** Human monocyte-derived macrophages

**Cell number:**  $3 \times 10^5$  cells per well

**Culture vessel:** 24-well plate

**Culture medium:** RPMI 1640 + 10% FBS + 1% Glutamax + 1% Sodium Pyruvate + 0.1%  $\beta$ -mercaptoethanol

**Nucleic Acid:** fluorescent oligonucleotide duplex (siGLO - Thermo Scientific)

Human macrophages were seeded and differentiated using 20ng/ml of M-CSF for 1 week in a 24-well plate. After differentiation, cells to be transfected with K2<sup>®</sup> Transfection Reagent were incubated with 5 $\mu$ l of K2<sup>®</sup> Multiplier in 600 $\mu$ l of culture medium and incubated for 2h.

Transfection solution was prepared in two separate vessels:

- 50  $\mu$ l of serum-free medium + desired amount of RNA oligo to achieve a final concentration of 50nM or 25nM (final volume of culture medium will be 600 $\mu$ l)
- 50  $\mu$ l of serum-free medium + 9  $\mu$ l or 4.5  $\mu$ l of transfection reagent

Transfection solutions were gently mixed, adding the nucleotide solution to the transfection reagent solution, and incubated for 20 min. Meanwhile, human macrophages were washed with PBS and 500  $\mu$ l of culture medium was added to each well.

Following that, 100 $\mu$ l of the transfection solution was added to each well, mixed by gently agitating the plate, and incubated for 24h.

After 24h cells were washed with PBS, fixed with 4% PFA for 15 min then the percentage of fluorescent cells was quantified by flow cytometry.

## Results

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Our results show a consistently higher effective transfection of our cells using the K2<sup>®</sup> Transfection system and visual observation also revealed a lower amount of dead cells.

